

## Cytology samples transportation guidelines

### Specialized guidelines according to the type of biological material:

#### ❖ Liquid-based cytology Test PAP

- ✓ Using a brush «Rovers Cervix-Brush», insert the core fibers of the brush in the endometrial canal.
- ✓ Maintaining steady and light pressure, turn the brush clockwise five times.
- ✓ Insert the brush in the liquid-based cytology vial. Remove the brush handle. Reseal the vial.
- ✓ Clearly fill out the label on the vessel so that the patient identification is possible.

#### ❖ Conventional cytology PAP test (PAP Smear)

- ✓ The patient should not use vaginal medication or vaginal lavages for 24 hours before taking the sample.
- ✓ The name of the patient should be marked with a black pencil on the rough side of the tile where the sample is to be placed.
- ✓ Insert the endoscope without using lubricant gel and collect the sample
- ✓ Any use of lubricant of any kind may render the sample useless.
- ✓ Using suitable spatulas and an endometrial brush, collect samples from the entire endo / exometrial surface. A number of suitable tools are now available to collect adequate endometrial specimens. Spatulae are sufficient for vaginal, ectopic and endometrial specimens.
- ✓ Quickly smear the collected material across the entire glass surface of the tile.

#### ❖ Body cavity fluids

- ✓ May include: pleural, ascetic, cystic or spinal fluids.
- ✓ Liquids that may coagulate must undergo anticoagulation (heparinisation) before they are submitted.
- ✓ Liquids can be transported to the laboratory in any suitable container with the relevant, complete and legible marking. Keep refrigerated until shipment.

#### ❖ Nipple secretions

- ✓ The few first drops of secretions should not be collected, as they mainly contain cellular debris.
- ✓ Using a tile in which the necessary marking has already been completed, slide it along the nipple to achieve a uniform spread.
- ✓ Fix immediately with cytology fixative in sprayable form.

- ✓ Repeat the procedure until the secretions stop.
- ✓ Aspiration of cystic fluid.
- ✓ Drain the liquid into a syringe and submit it to our lab, with the name of the patient on a label.
- ✓ Until they are shipped to the laboratory, samples should be kept refrigerated.

❖ **Solid mass aspiration**

- ✓ Absorb the sample with a syringe
- ✓ Label the syringe with the name of the patient
- ✓ Submit it to our laboratory or store under refrigeration until you submit it.

❖ **Sample collection with a protected bronchial brush – PBB**

- ✓ Samples that have been collected using a protected bronchial brush can be submitted in two different ways:
  1. 1. Place the brush in a test tube with a screw cap filled with saline and submit it to our laboratory immediately.
  2. 2. Apply the sample from the brush to a glass tile in which you have previously filled the label (use pencil). Ensure that the entire sample is spread evenly on the tile.

❖ **BrochoAlveolar Lavage – BAL**

- ✓ The samples should be sent inside a clean, airtight and securely sealed tube or container.
- ✓ The samples should be transported to the laboratory as soon as possible in a container with a fully filled out and legible label. If immediate transfer is not feasible, store the specimen under cooling or place it in a bowl filled with "CytoRich Red" to avoid spoilage of the sample cells.

❖ **Sputum samples**

- ✓ The only acceptable samples are those produced from expectoration after deep coughing. Saliva and samples infected with nasal discharge or food are not acceptable.
- ✓ Advise the patient to collect the sample in the morning, before anything else. Ask him to take a deep breath and spit out the mucus that will be caused by the deep cough in a suitable pot that you will have provided him.
- ✓ If patients cannot produce any sputum out of their lungs, consider whether the sputum induction process is possible in the first place.

- ✓ The samples should be transported to the laboratory as soon as possible in a container with a fully filled out and legible label. If immediate transfer is not feasible, store the specimen under cooling or place it in a bowl filled with "CytoRich Red" to avoid spoilage of the sample cells.

❖ <sup>7</sup> For small amounts of liquid (1  
**Thyroid gland liquid sample collection**

- ✓ Γα -2ml), you can smear the sample on a glass laboratory tile. Place a second tile on the first and remove it to create a uniform spread. Spray immediately with a fixative.
- ✓ For Wright stain tiles: Repeat the staining procedures for one or two tiles and allow to dry in the air. Please indicate on the tile that it has dried in the air.
- ✓ For larger quantities of liquid: Send the sample to a sealed syringe for processing. Refrigerate the sample if immediate shipment is not possible.

❖ **Tzank stain collection procedure (for herpes)**

- ✓ Open the rash from which the sample will be collected with a sterile blade or biology needle.
- ✓ Scrape the base of the rash with a tongue depressor, a spatula or a sterile blade. If there is liquid, you can collect it with a biology needle.
- ✓ Immediately smear the specimen onto a glass tile on which you have already marked the patient's name using a pencil.
- ✓ Spray the tile immediately with a cytology fixative. If you do not, then the sample will air-dry, and the diagnosis will not be possible.
- ✓ Place the samples in a plastic container used for Pap tests. Send it to the lab.

❖ **Urine**

- ✓ Samples not coming from the first morning urine are preferable
- ✓ Minimum sample volume: 5ml. 50 to 100 ml are preferable.
- ✓ Samples collected by catheter or from the first morning urine should be transferred to a clean container with a legible and completely filled-out label.
- ✓ All specimens should be transported as soon as possible to the laboratory or, if that is not possible, stored under refrigeration to avoid cellular degeneration. Samples from a 24-hour collection are not accepted.